日本農芸化学会2018年度大会 SCIEX協賛ランチョンセミナー

過酸化脂質(脂質ヒドロペルオキシド)の質量分析



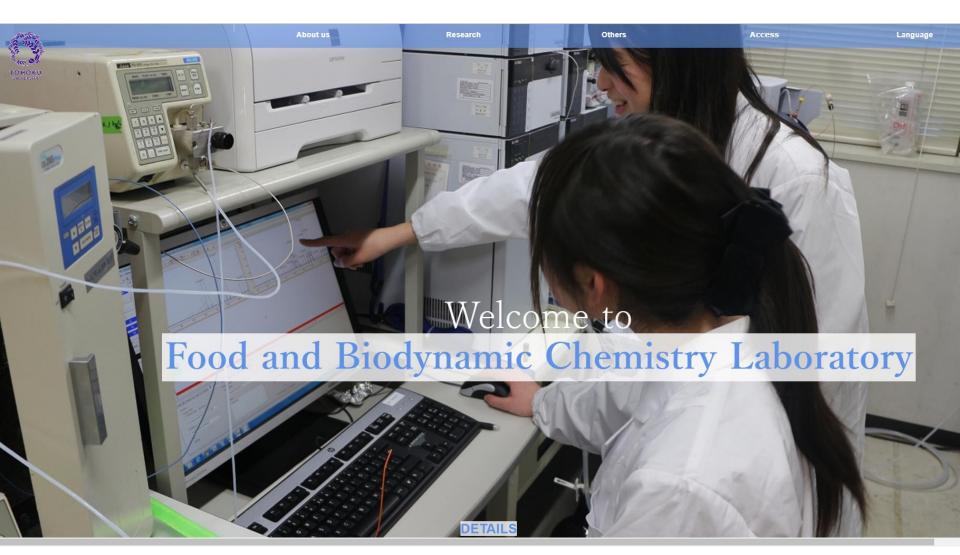






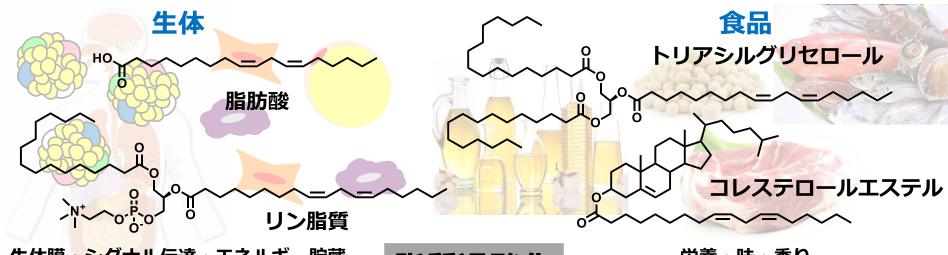


Food & Biodynamic Chemistry Laboratory



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脂質過酸化は生体老化や食品劣化に大きく関与する

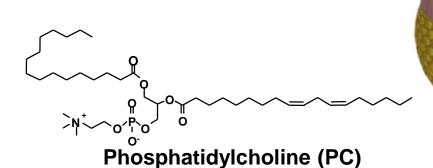


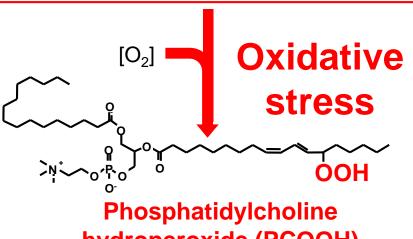
生体膜・シグナル伝達・エネルギー貯蔵

脂質過酸化

栄養・味・香り

What's PCOOH?





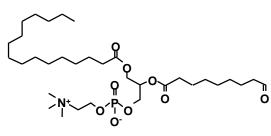
hydroperoxide (PCOOH)



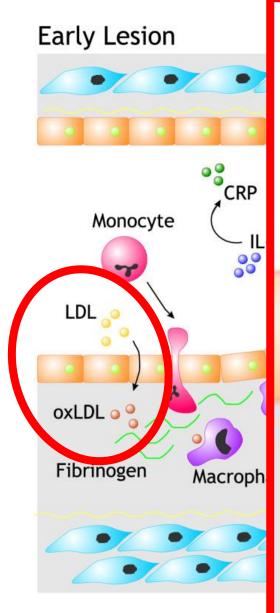
Chemiluminescence detection (CL)-HPLC

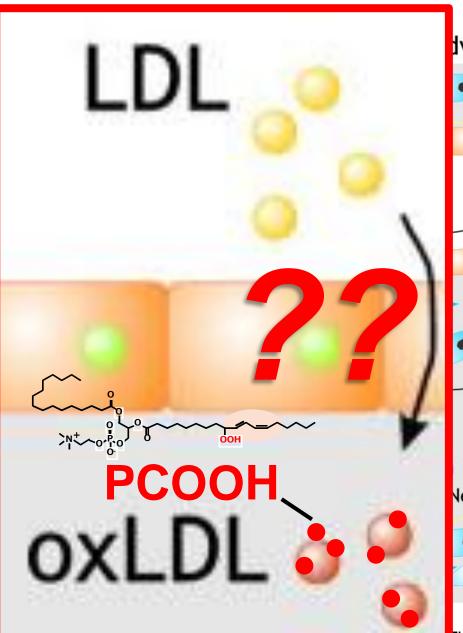
T. Miyazawa et al. J Lipid Res. 1992

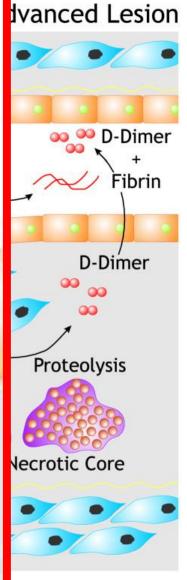
Phosphatidylcholine hydroxide (PCOH)



sn2-Truncated phosphatidylcholine

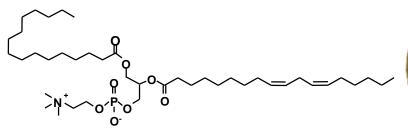




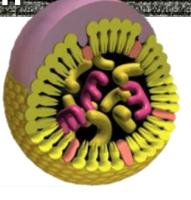


Coronic Ct a.., Thrombosis Journal 2004

Phospholipid peroxidation









✓ Enzymatic oxidation

✓ Auto-oxidation

✓ Singlet oxygen-induced oxidation

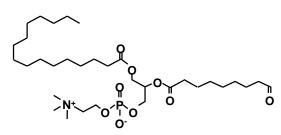
A Singlet OOH

 $[O_2]$

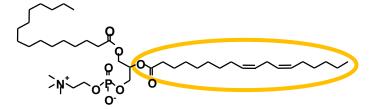
Phosphatidylcholine hydroperoxide (PCOOH)

PC is oxidized by three distinct mechanisms.

Phosphatidylcholine hydroxide (PCOH)



sn2-Truncated phosphatidylcholine



PC possessing linoleic acid at sn-2

(Singlet oxygen-induced oxidation)

HpODE: Hydroperoxyoctadecadienoic

(Singlet oxygen-induced oxidation)

16:0/13-HpODE PC

(Singlet oxygen-induced oxidation Auto-oxidation, Enzymatic oxidation)

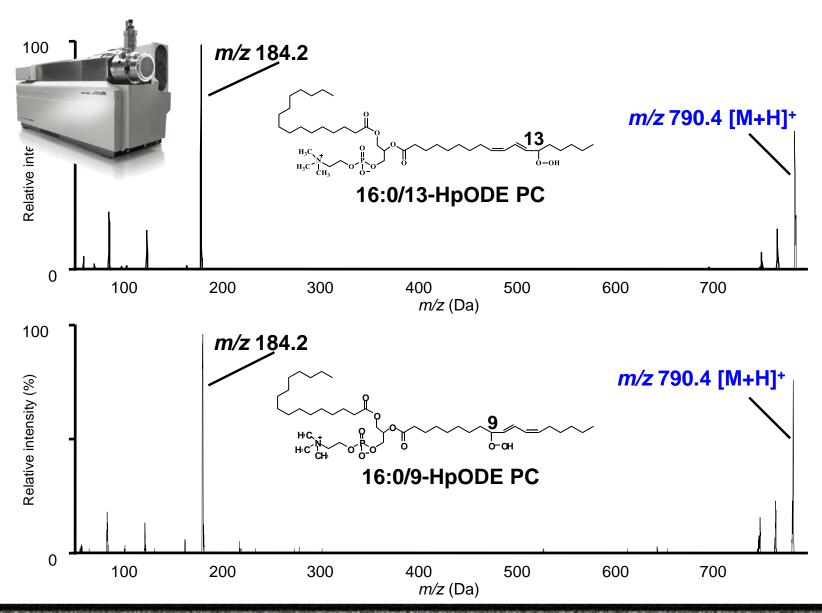
OOH OOH

16:0/9-HpODE PC

(Singlet oxygen-induced oxidation, Auto-oxidation, Enzymatic oxidation)

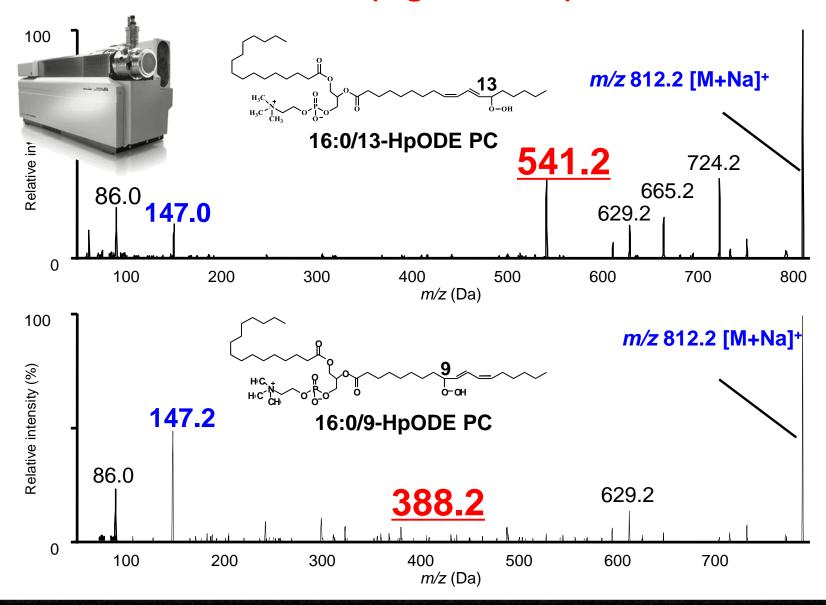
Product ion mass spectra (MS/MS) of 16:0/HpODE PC

Absence of alkali metals

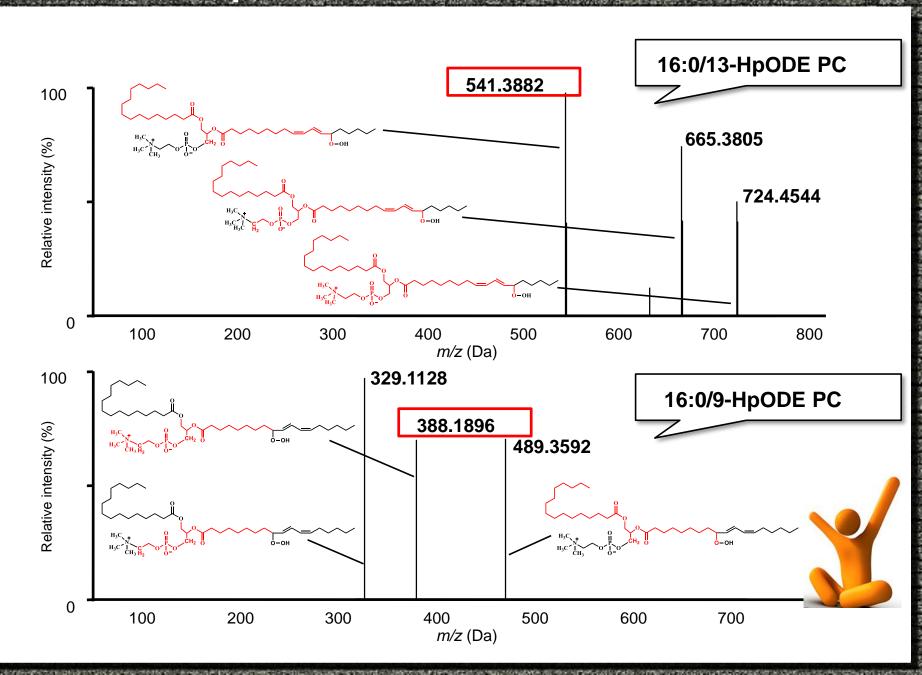


Product ion mass spectra (MS/MS) of 16:0/HpODE PC

Presence of alkali metals (e.g., sodium)

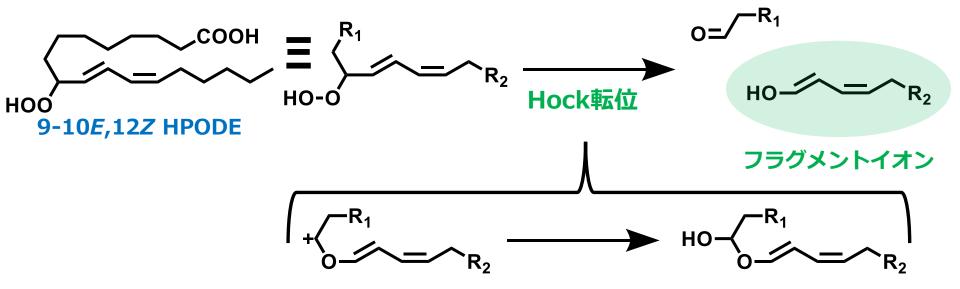


Differences of product ion intensities



a開裂

Hock転位





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Published online: 30 August 2017

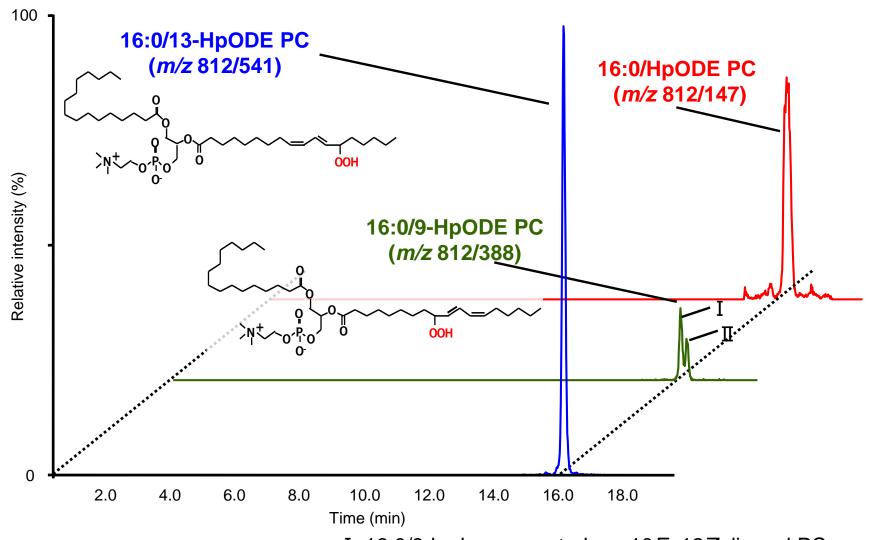
OPEN A novel chiral stationary phase LC-MS/MS method to evaluate oxidation mechanisms of edible oils

Junya Ito¹, Naoki Shimizu¹, Eri Kobayashi¹, Yasuhiko Hanzawa¹, Yurika Otoki¹, Shunji Kato^{1,2}, Takafumi Hirokawa³, Shigefumi Kuwahara³, Teruo Miyazawa^{1,4} & Kiyotaka Nakagawa¹

The elucidation of lipid oxidation mechanisms of food is vital. In certain lipids, characteristic lipid hydroperoxide isomers are formed by different oxidation mechanisms (i.e., photo-oxidation or autooxidation). For example, linoleic acid is photo-oxidized to 13-9Z, 11E-hydroperoxyoctadecadienoic acid (HPODE), 12-9Z,13E-HPODE, 10-8E,12Z-HPODE and 9-10E,12Z-HPODE, whereas 13-9Z, 11E-HPODE, 13-9E,11E-HPODE, 9-10E,12Z-HPODE and 9-10E,12E-HPODE are formed by auto-oxidation. Therefore, we considered that oxidation mechanisms could be evaluated by analyzing these characteristic positional and cis/trans lipid hydroperoxide isomers. In this study, we developed a novel chiral stationary phase LC-MS/MS (CSP-LC-MS/MS) method to analyze the positional and cis/trans isomers of HPODE, with the use of a chiral column and sodium ion. Also, as an application of the method, either light-exposed or heated edible oils were treated with lipase to hydrolyze triacylglycerols. The resultant fatty acids including HPODE isomers were analyzed with the developed method. As a result, HPODE isomers characteristic to photo-oxidation were certainly detected in light-exposed edible oils. On the other hand, in heated edible oils, the HPODE isomers characteristic to auto-oxidation were largely increased. Thus, the combination of the developed CSP-LC-MS/MS method with lipase proves to be a powerful tool to evaluate the involvement and mechanisms of lipid oxidation in the process of food deterioration.

Measurement of PCOOH in clinical specimens

Plasma PCOOH in patients with hyperlipidemia



I .16:0/9-hydroperxyoctadeca-10*E*, 12*Z*-dienoyl PC II .16:0/9-hydroperxyoctadeca-10*E*, 12*E*-dienoyl PC

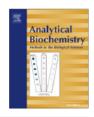
HPLC-MS/MS with MRM



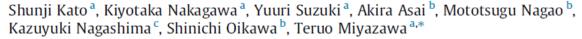
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Liquid chromatography-tandem mass spectrometry determination of human plasma 1-palmitoyl-2-hydroperoxyoctadecadienoyl-phosphatidylcholine isomers via promotion of sodium adduct formation



^a Food and Biodynamic Chemistry Laboratory, Graduate School of Agricultural Science, Tohoku University, Sendai 981-8555, Japan







Article

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Tandem Mass Spectrometry Analysis of Linoleic and Arachidonic Acid Hydroperoxides via Promotion of Alkali Metal Adduct Formation

Junya Ito, Shunsuke Mizuochi, Kiyotaka Nakagawa,* Shunji Kato, and Teruo Miyazawa

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b Division of Endocrinology and Metabolism, Department of Medicine, Nippon Medical School, Tokyo 113-8603, Japan

c Cardiovascular Institute, Tokyo 106-0031, Japan

Measurement of PCOOH in clinical specimens

	16:0/13-HpODE PC	16:0/9-HpODE PC	16:0/HpODE PC*		
		pmol/mL			
Healthy subjects n=8	25.4	22.6	49.8		
	46.3	41.0	86.4		
	50.5	46.0	103.4		
	34.7	34.3	67.3		
	31.9	30.9	66.1		
	49.3	43.5	102.7		
	19.0	16.6	38.2		
	31.9	30.3	64.8		
	86.9	66.0	138.6		
	32.9	32.2	64.4		
	45.5	35.0	90.3		
	39.0	36.0	77.8		
Patients with hyperlipidemia n=12	42.8	38.9	87.2		
	112.5	84.3	194.6		
	49.8	50.1	99.5		
	67.4	69.4	132.5		
	33.3	30.2	61.9		
	43.6	34.9	80.6		
	32.3	32.9	61.3		
	42.9	32.8	79.1		

16:0/13-HpODE PC + 16:0/9-HpODE PC = 16:0/HpODE PC

Discussion

16:0/13-HpODE PC + 16:0/9-HpODE PC = 16:0/HpODE PC

$$\begin{array}{c} & & & & & & & \\ & & & & & \\ & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\$$

16:0/13-HpODE PC

(Auto-oxidation, Enzymatic oxidation, Singlet oxygen induced exidation

16:0/9-HpODE PC

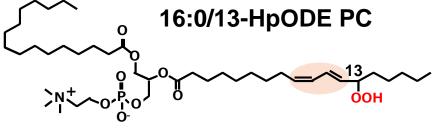
(Auto-oxidation, Enzymatic oxidation, Singlet exygen induced exidation

Auto-oxidation and/or enzymatic oxidation, rather than singlet oxygen-induced oxidation, would cause PC peroxidation.

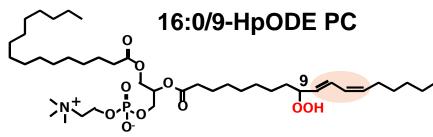
Discussion & on-going study

16:0/10-HpODE PC (Singlet oxygen-induced oxidation)

16:0/12-HpODE PC (Singlet oxygen-induced oxidation)

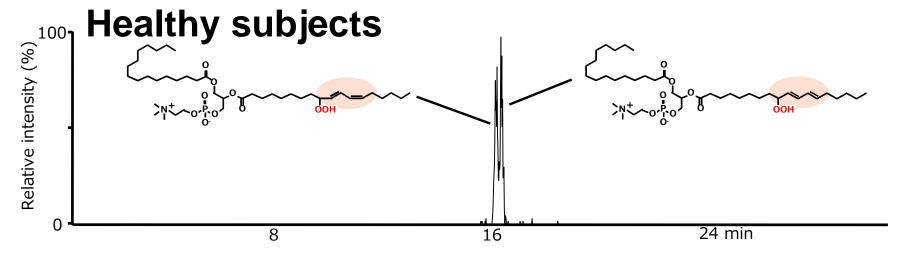


(Auto-oxidation, Enzymatic oxidation, Singlet exygen induced exidation)

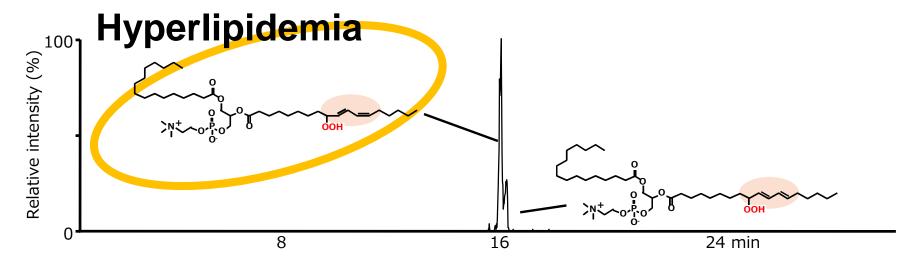


(Auto-oxidation, Enzymatic oxidation, Singlet-exygen induced exidation)

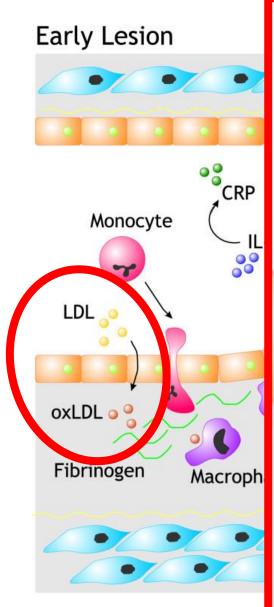
On-going study

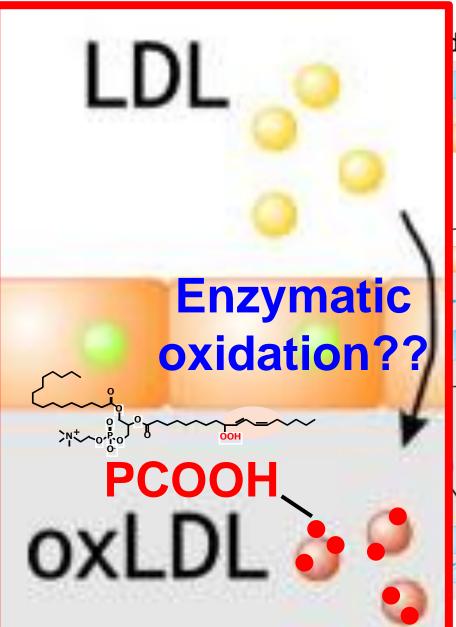


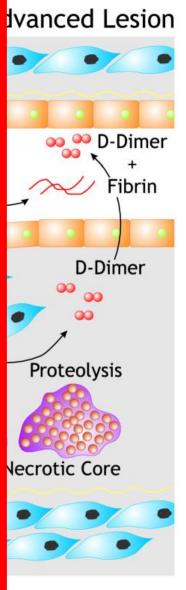
Auto-oxidation, Enzymatic oxidation



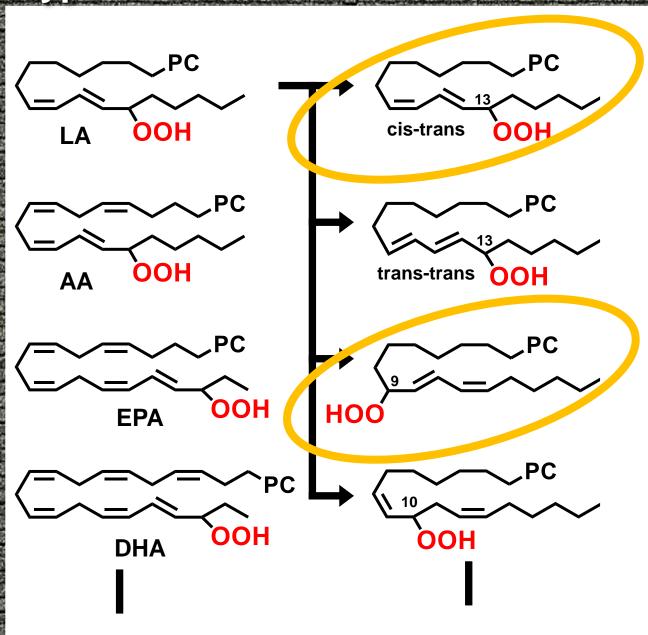
Auto-oxidation, Enzymatic oxidation

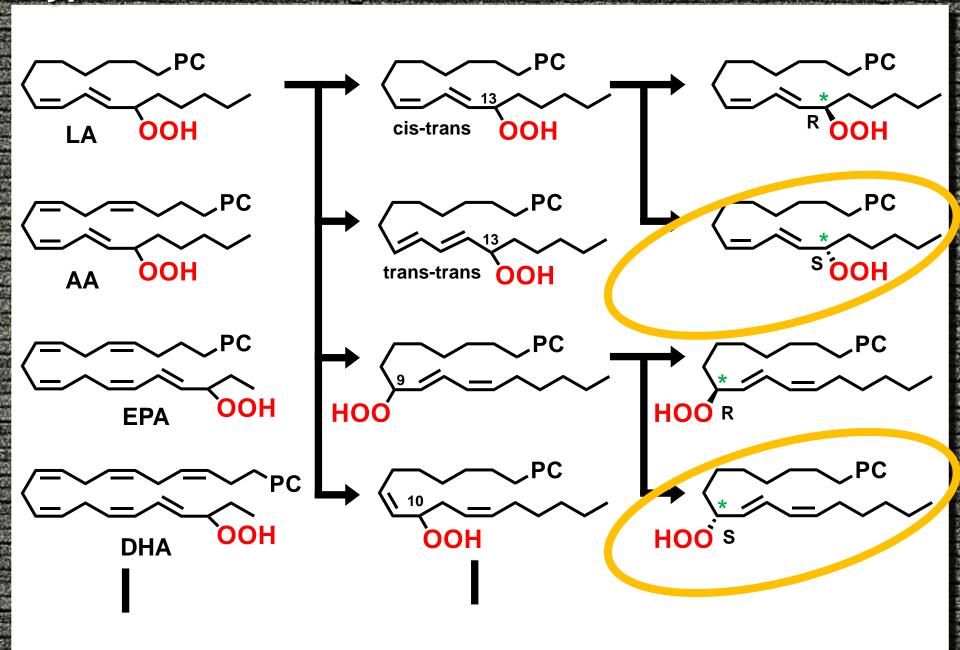






Coronic Ct al., Thrombosis Journal 2004

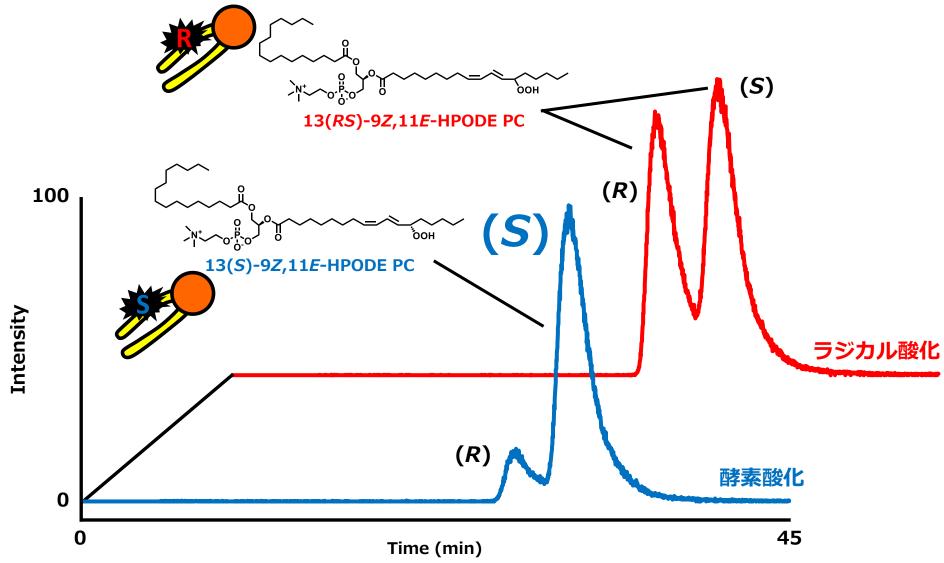




立体異性分離(RS分離)可能な条件を見出した

Chiral column	Condition 1	Condition 2	Condition 3	Condition 4	Condition 5	Condition 6	Condition 7	Condition 8	Condition 9
	Rs t1 t2								
IA	0.0 30 30	0.0 22 22	0.0 17 17	0.0 16 16	nd nd nd	0.0 16 16	0.0 3 3	0.0 2 2	not-tested
IB	0.0 30 30	0.0 25 25	0.0 27 27	0.0 25 25	nd nd nd	nd nd nd	0.0 5 5	0.0 2 2	not-tested
IC	0.0 30 30	0.0 27 27	0.0 7 7	0.0 7 7	nd nd nd	nd nd nd	0.0 8 8	0.0 3 3	not-tested
ID	0.0 16 16	0.0 14 14	0.0 6 6	0.0 6 6	nd nd nd	0.0 15 15	0.0 3 3	0.0 2 2	not-tested
IE	nd nd nd	nd nd nd	0.0 15 15	0.0 16 16	nd nd nd	nd nd nd	0.0 8 8	0.0 3 3	not-tested
IF	nd nd nd	nd nd nd	0.0 30 30	0.0 27 27	nd nd nd	nd nd nd	0.0 5 5	0.0 3 3	not-tested
AD-RH	0.0 12 12	0.0 6 6	0.0 5 5	0.0 51 51	not-tested	not-tested	not-tested	not-tested	not-tested
AS-RH	nd nd nd	nd nd nd	0.0 5 5	0.0 5 5	not-tested	not-tested	not-tested	not-tested	not-tested
AY-RH	nd nd nd	0.0 8 8	0.0 6 6	0.0 6 6	not-tested	not-tested	not-tested	not-tested	not-tested
OD-RH	0.0 18 18	0.0 15 15	0.0 9 9	0.0 9 9	not-tested	not-tested	not-tested	not-tested	not-tested
OJ-RH	nd nd nd	nd nd nd	0.0 5 5	0.0 5 5	not-tested	not-tested	not-tested	not-tested	not-tested
OZ-RH	0.0 3 3	nd nd nd	0.0 13 13	0.0 13 13	not-tested	not-tested	not-tested	not-tested	not-tested
AD-H	not-tested	not-tested	not-tested	not-tested	0.0 35 35	0.0 6 6	0.0 2 2	0.0 2 2	not-tested
АҮ-Н	not-tested	not-tested	not-tested	not-tested	0.0 11 11	0.0 8 8	0.0 2 2	0.0 2 2	not-tested
OD-H	not-tested	not-tested	not-tested	not-tested	0.0 18 18	0.0 5 5	0.0 2 2	0.0 2 2	not-tested
ОЈ-Н	not-tested	not-tested	not-tested	not-tested	0.0 6 6	0.0 6 6	0.0 2 2	0.0 1 1	not-tested
OZ-H	not-tested	not-tested	not-tested	not-tested	0.0 7 7	0.0 8 8	0.0 3 3	0.0 3 3	not-tested
OP (+)	not-tested	1.6 24 27							

Rs(分離度)≧1.5で完全分離; t1, 第1ピーク溶出時間; t2, 第2ピーク溶出時間



J. Ito, et al., *J. Chromatogr. A*, 1386:53-61 (2015)

酵素酸化とラジカル酸化・一重項酸素酸化を明確に区別できる



Contents lists available at ScienceDirect

Journal of Chromatography A

journal homepage: www.elsevier.com/locate/chroma



Direct separation of the diastereomers of phosphatidylcholine hydroperoxide bearing 13-hydroperoxy-9*Z*,11*E*-octadecadienoic acid using chiral stationary phase high-performance liquid chromatography



Junya Ito^a, Kiyotaka Nakagawa^{a,*}, Shunji Kato^a, Takafumi Hirokawa^b, Shigefumi Kuwahara^b, Toshiharu Nagai^c, Teruo Miyazawa^a

- ³ Food and Biodynamic Chemistry Laboratory, Graduate School of Agricultural Science, Tohoku University, Sendai, 981-8555, Japan
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Keywords:
Diastereomer separation
Chiral stationary phase
Diastereomer
HPLC-Q-TOF/MS
Phosphatidylcholine hydroperoxide

ABSTRACT

Increasing evidence suggests that phospholipid peroxidation plays important roles in the pathogenesis of various diseases, such as atherosclerosis. With regard to the biochemical processes that initiate phospholipid peroxidation in vivo, enzymatic conversion of phosphatidylcholine to phosphatidylcholine hydroperoxide (PCOOH) by lipoxygenase (LOX) may play a crucial role. This will become clear if we can analyze PCOOH bearing hydroperoxy fatty acids with S-stereoconfiguration. In this study, we therefore attempted such an analysis. Initially, we used LOX, linoleic acid and Lyso phosphatidylcholine, and synthesized PCOOH bearing 13S-hydroperoxy-9Z,11E-octadecadienoic acid (13(S)-9Z,11E-HPODE). PCOOH bearing racemic 13-9Z,11E-HPODE was also prepared. We used liquid chromatography equipped with CHIRALPAK OP (+) (poly (o-pyridyl diphenylmethacrylate) coated on silica), a UV detector and a quadrupole-time-of-flight mass spectrometer, and achieved diastereomer separation of PCOOH stereoisomers with excellent resolution and peak shape. This is the first study reporting the diastereomer separation of PCOOH. The present method will be beneficial in developing a better understanding of the biochemical processes that initiate oxidative stress (PCOOH formation) in vivo, which may lead to further elucidation of the involvement of PCOOH in the development of diseases. In addition to clinical applications, the present method may also be effective in the evaluation of enzymatic oxidative food deterioration.

^c Tsukishima Foods Industry Co., Ltd., Tokyo, 134-8520, Japan

CrossMark

RESEARCH PAPER

A novel chiral stationary phase HPLC-MS/MS method to discriminate between enzymatic oxidation and auto-oxidation of phosphatidylcholine

Junya Ito¹ · Kiyotaka Nakagawa¹ · Shunji Kato¹ · Takafumi Hirokawa² · Shigefumi Kuwahara² · Toshiharu Nagai³ · Teruo Miyazawa^{1,4}

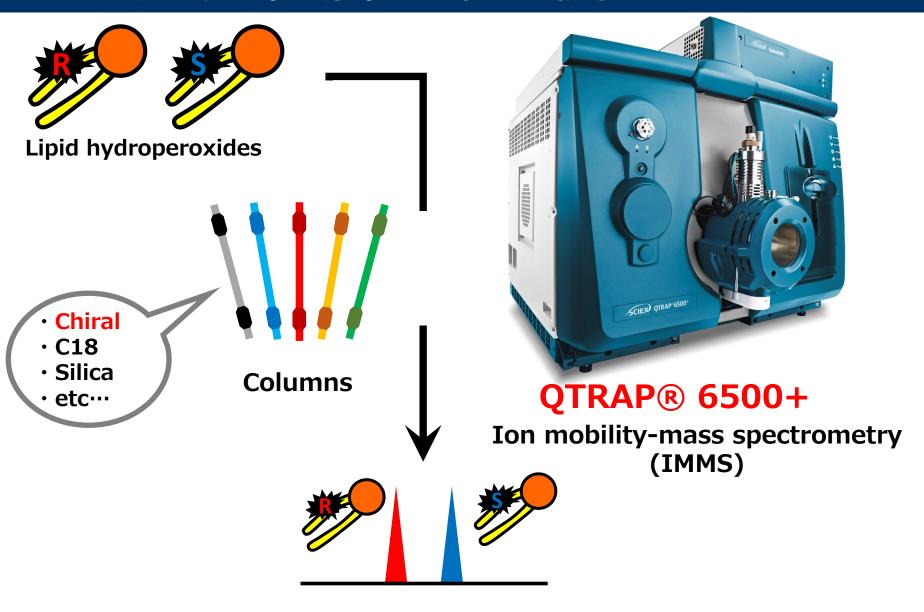
Received: 22 May 2016 / Revised: 6 August 2016 / Accepted: 15 August 2016 / Published online: 22 August 2016 © Springer-Verlag Berlin Heidelberg 2016

Abstract To elucidate the role of enzymatic lipid peroxidation in disease pathogenesis and in food deterioration, we recently achieved stereoselective analysis of phosphatidylcholine hydroperoxide (PCOOH) possessing 13S-hydroperoxy-9Z,11E-octadecadienoic acid (13(S)-9Z,11E-HPODE) using HPLC-MS/MS with a CHIRALPAK OP (+) column. Because enzymatic oxidation progresses concurrently with auto-oxidation, we need to distinguish them further. Here, we attempted such an analysis. First, we used lipoxygenase, linoleic acid, and lysophosphatidylcholine (LPC) to synthesize the enzymatic oxidation product 13(S)-9Z,11E-HPODE PC, and the auto-oxidation products 13(RS)-9Z,11E-HPODE PC and 13(RS)-9E,11E-HPODE PC, which were used as standards to test the ability of various columns to separate the enzymatic oxidation product from auto-oxidation products. Separation was achieved by connecting in series two columns with different properties: CHIRALPAK OP (+) and CHIRALPAK IB-3. The CHIRALPAK OP (+) column separated 13(*R*)-9*Z*,11*E*-HPODE PC and 13(*S*)-9*Z*,11*E*-HPODE PC, whereas CHIRALPAK IB-3 enabled separation of 13(*S*)-9*Z*,11*E*-HPODE PC and 13(*RS*)-9*E*,11*E*-HPODE PC. The results for the analysis of both enzymatically oxidized and auto-oxidized lecithin (an important phospholipid mixture in vivo and in food) indicate that our method would be useful for distinguishing enzymatic oxidation and auto-oxidation reactions. Such information will be invaluable for elucidating the involvement of PCOOH in disease pathogenesis and in food deterioration.

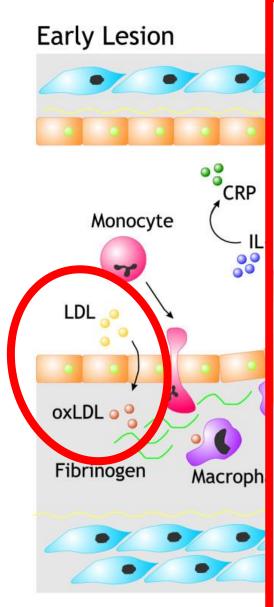
Keywords Chiral stationary phase · Chiral column · Phosphatidylcholine hydroperoxide · Lipid oxidation · LC-MS/MS

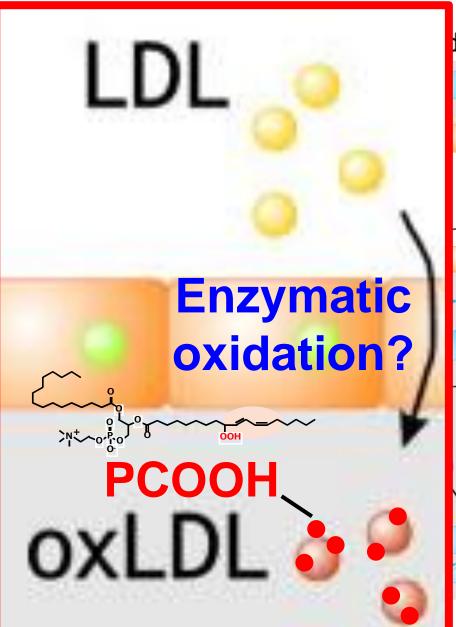
Abbreviations

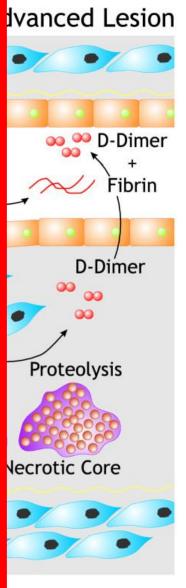
16:0 LPC 1-palmitoyl-2-hydroxy-sn-glycero-3-



LC-IMS-MS/MS法を構築する

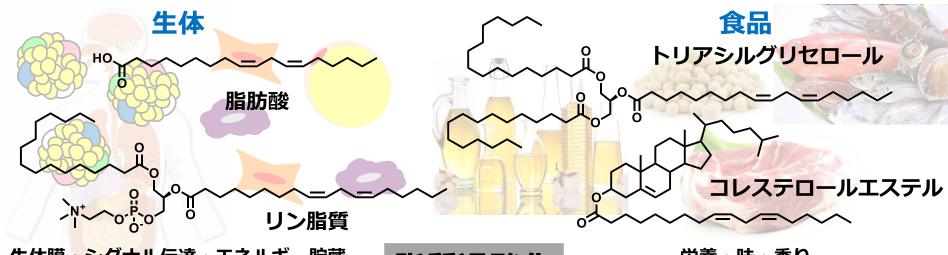






Coronic Ct a.., Thrombosis Journal 2004

脂質過酸化は生体老化や食品劣化に大きく関与する

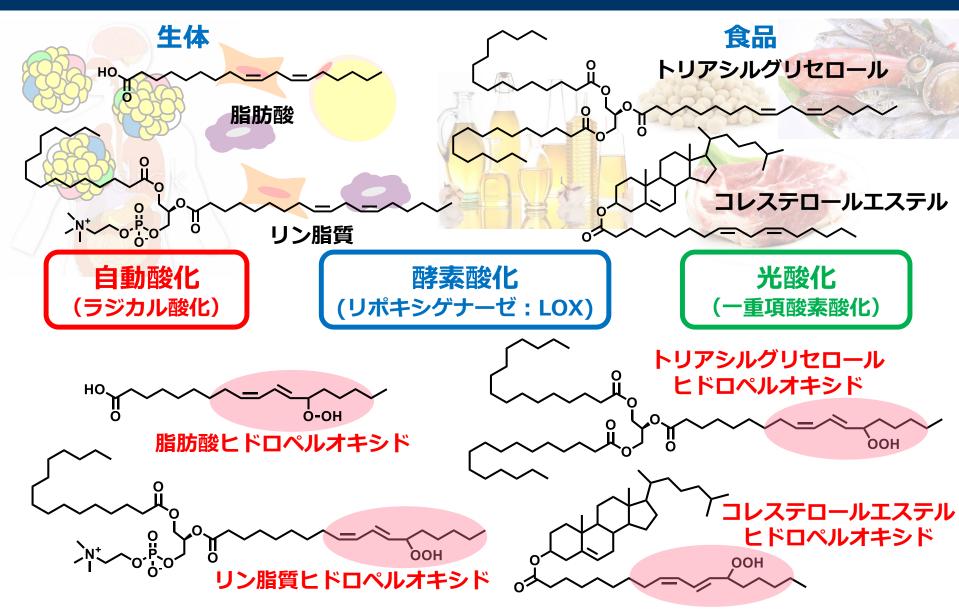


生体膜・シグナル伝達・エネルギー貯蔵

脂質過酸化

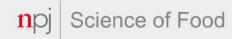
栄養・味・香り

まとめ



生体や食品の脂質過酸化機構を解明し、生体老化や食品劣化を予防する





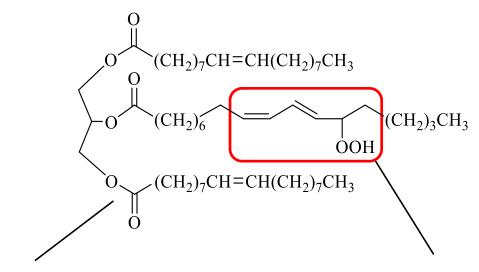


Determination of triacylglycerol oxidation mechanisms in canola oil using liquid chromatography-tandem mass spectrometry

Shunji Kato^{1,2}, Naoki Shimizu¹, Yasuhiko Hanzawa¹, Yurika Otoki¹, Junya Ito¹, Fumiko Kimura³, Susumu Takekoshi², Masayoshi Sakaino⁴, Takashi Sano⁴, Takahiro Eitsuka¹, Teruo Miyazawa^{5,6}, Kiyotaka Nakagawa^{1*}

Triacylglycerol (TG), the main component of edible oil, is oxidized by thermal- or photo- oxidation to form TG hydroperoxide (TGOOH) as the primary oxidation product. Since TGOOH and its subsequent oxidation products cause not only the deterioration of oil quality but also various toxicities, preventing the oxidation of edible oils is essential. Thereby understanding oxidation mechanisms that cause the formation of TGOOH is necessary. Since isomeric information of lipid hydroperoxide provides insights about oil oxidation mechanisms, we focused on dioleoyl-(hydroperoxy octadecadienoyl)-TG (OO-HpODE-TG) isomers, which are the primary oxidation products of the most abundant TG molecular species (dioleoyl-linoleoyl-TG) in canola oil. To secure highly selective and sensitive analysis, authentic OO-HpODE-TG isomer references (i.e. hydroperoxide positional/geometrical isomers) were synthesized and analyzed with HPLC-MS/MS. With the use of the method, photo- or thermal- oxidized edible oils were analyzed. While dioleoyl-(10-hydroperoxy-8E,12Z-octadecadienoyl)-TG (OO-(10-HpODE)-TG) and dioleoyl-(12-hydroperoxy-9Z,13E-octadecadienoyl)-TG (OO-(12-HpODE)-TG) were characteristically detected in photo-oxidized oils, dioleoyl-(9hydroperoxy-10E,12E-octadecadienoyl)-TG and dioleoyl-(13-hydroperoxy-9E,11E-octadecadienoyl)-TG were found to increase depending on temperature in thermal-oxidized oils. These results prove that our methods not only evaluate oil oxidation in levels that are unquantifiable with peroxide value, but also allows for the determination of oil oxidation mechanisms. From the analysis of marketed canola oils, photo-oxidized products (i.e. OO-(10-HpODE)-TG and OO-(12-HpODE)-TG) were characteristically accumulated compared to the oil analyzed immediately after production. The method described in this paper is valuable in the understanding of oil and food oxidation mechanisms, and may be applied to the development of preventive methods against food deterioration.

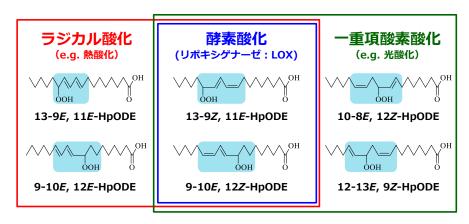
TGOOH解析に必要な情報



①構成脂肪酸

オレイン酸、リノール酸 リノレン酸、EPA、DHA… 組み合わせ

②酸素結合位置(異性体)



定量性と汎用性を考慮した2つの方法を構築

TGOOH標品の調製

リノール酸

酸化 (ラジカル+光酸化) 精製(分取HPLC)

リノール酸ヒドロペルオキシド (HpODE)異性体

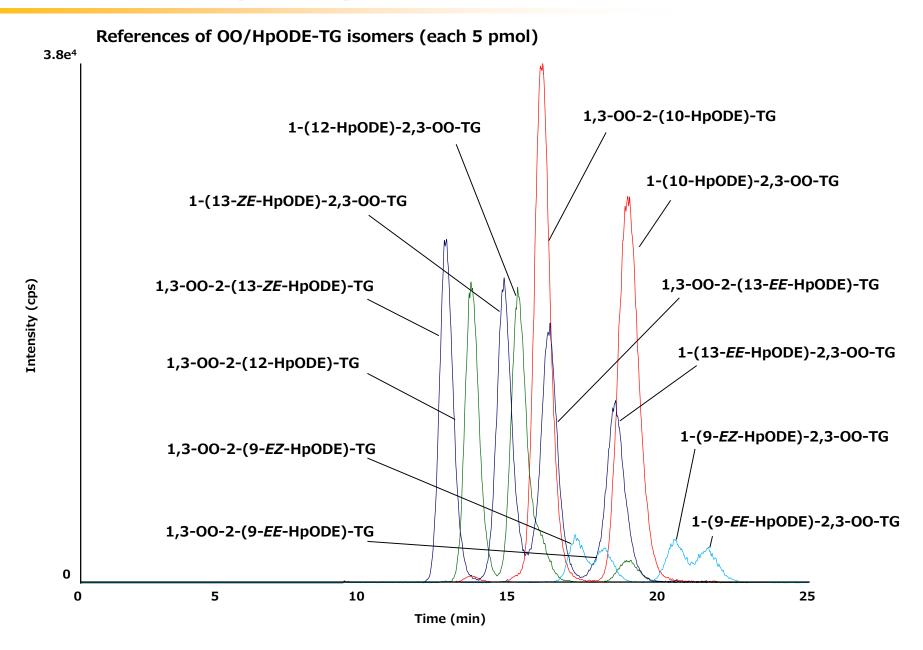
> ヒドロペルオキシド基の保護 (PPTS, MxP)

LAOOMXP

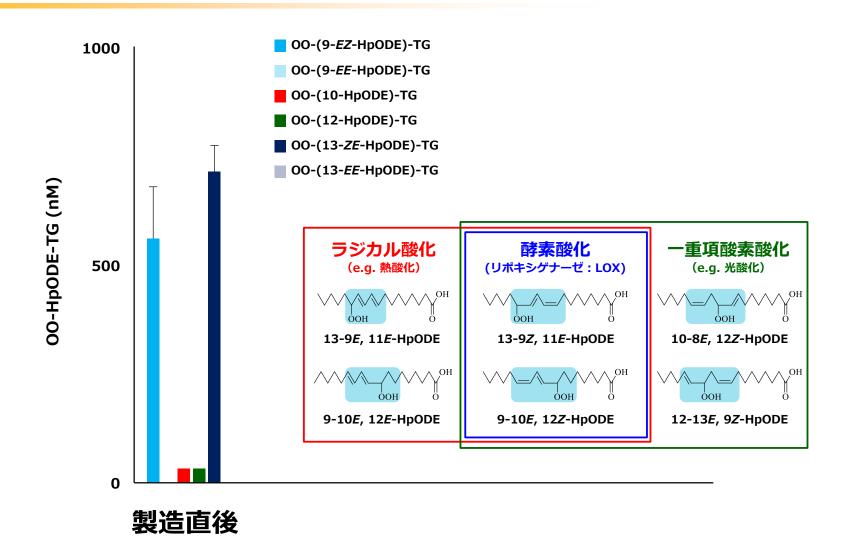
エステル化 Dioleoyl glycerol, DCC, DMAP
脱保護

OOL/OLO type TGOOH isomers 9-hydroperxy-octadeca-10*E*, 12*Z*-dienoic acid 9-hydroperxy-octadeca-10*E*, 12*E*-dienoic acid 10-hydroperxy-octadeca-8*E*, 12*Z*-dienoic acid 12-hydroperxy-octadeca-13*E*, 9*Z*-dienoic acid 13-hydroperxy-octadeca-9*E*, 11*E*-dienoic acid 13-hydroperxy-octadeca-9*Z*, 11*E*-dienoic acid

TGOOH異性体 (標準品)

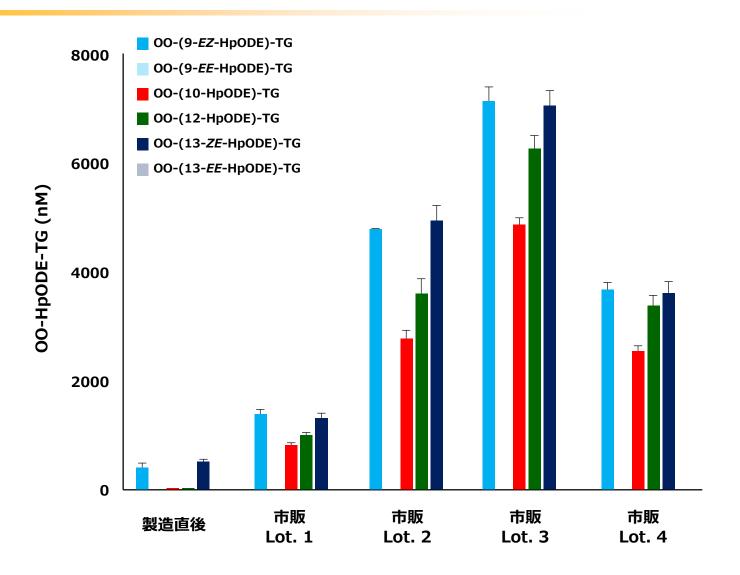


サンプル解析



新鮮な油脂(POV=0)に含まれるTGOOHも 高感度に定量が可能

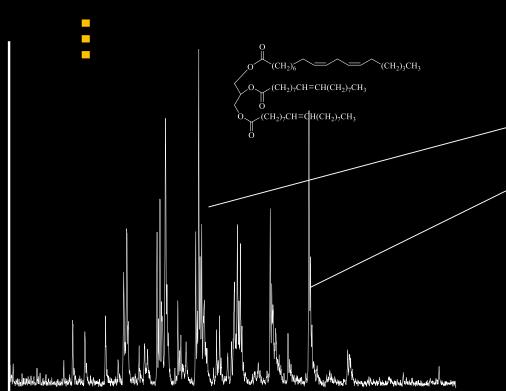
サンプル解析



店頭に並んでいる間は主に光酸化が亢進

様々な油脂へと応用

Olive oil
Fish oil
Rice bran oil
Sesame oil







Oleic acid a-linolenic acid y-linolenic acid DHA



